



Bioinformatic Command-line tools

sol genomics network
Boyce Thompson Institute for Plant
Research
Ithaca, New York 14853
U.S.A.

presented by
Naama Menda



Command-line

- Linux based
- More options running programs
- Run programs locally
- Run jobs on bigger server





Command-line tools

- Local BLAST
- RNAseq analysis
 - Pre-processing (FastQC, fastq-mcf)
 - Assembly (Bowtie, Tophat, Trinity)
 - View output (samtools tview)
 - Expression (Cufflinks)





Reference-guided Assembly



TopHat
Aligns RNA-Seq reads to the genome using Bowtie
Discovers splice sites

- ◆ <http://tophat.cbcb.umd.edu/>
- ◆ deals with splice junctions
- ◆ Illumina RNA-Seq reads (paired or single end)





Exercise 1

Two RNA-seq datasets used in the tomato genome project were downloaded from the SRA in .sra format and extracted using the SRA toolkit (NCBI).

<http://www.ncbi.nlm.nih.gov/sra>

They were cleaned using fastq-mcf in last week's class. All data is from *S. pimpinellifolium*.

Datasets :

- breaker fruit (two files)
- immature fruit (two files)

In this exercise, we will map the reads to tomato chromosome 4 using reference-guided assembly





- Use bowtie2-build to index the reference
- Index tomato reference file
 - *bowtie2-build SL2.40ch04.fa SL2.40ch04*
- * Download files from
ftp.solgenomics.net/bioinfo_class/interns/data/





- Use tophat2 to map each .fq read set to the reference

```
tophat2 --no-coverage-search  
--no-novel-juncs -o SRR404334_tophat_out/  
~/bwt2_index/SL2.40ch04 SRR404334_ch4.fq
```

Repeat tophat2 command for remaining 3 data sets
(1 more breaker, 2 immature fruit).



Tophat output



sol genomics network

Tophat output:

- ◆ bam file of mapped reads (bam = compressed .sam file)
- ◆ bed files: junctions, insertions, deletions

Convert accepted_hits.bam to accepted_hits.sam

samtools view -o accepted_hits.sam accepted_hits.bam



* reference-guided assembly output

Response	Percentage
Good job	75%
Not a good job	25%



Sam format column definitions

Index	Field Name	Description
1	QNAME	Query pair NAME if paired; or Query NAME if unpaired
2	FLAG	Bitwise FLAG
3	RNAME	Reference sequence NAME
4	POS	1-based leftmost POSition of the clipped sequence
5	MAPQ	MAPping Quality
6	CIGAR	Extended CIGAR string
7	MRNM	Mate Reference sequence NaMe; "=" if the same as RNAME
8	MPOS	1-based leftmost Mate POSition of the clipped sequence
9	ISIZE	Inferred Insert SIZE
10	SEQ	Query SEQUENCE
11	QUAL	Query QUALity

Table 1.1: Brief summary of the SAM format



Sam flag field definitions

Field	Hex Code	Description
f_0	0x0001	the read is paired in sequencing, no matter whether it is mapped in a pair
f_1	0x0002	the read is mapped in a proper pair (depends on the protocol, normally inferred during alignment)
f_2	0x0004	the query sequence itself is unmapped
f_3	0x0008	the mate is unmapped
f_4	0x0010	strand of the query (0 for forward; 1 for reverse strand)
f_5	0x0020	strand of the mate
f_6	0x0040	the read is the first read in a pair
f_7	0x0080	the read is the second read in a pair
f_8	0x0100	the alignment is not primary (a read having split hits may have multiple primary alignment records)
f_9	0x0200	the read fails platform/vendor quality checks
f_{10}	0x0400	the read is either a PCR duplicate or an optical duplicate

Table 1.2: Brief summary of the SAM format

Excellent explanation for bitwise flags found here:
<http://seqanswers.com/forums/showthread.php?t=2301>

Tool to translate meaning of bitwise flag:
<http://picard.sourceforge.net/explain-flags.html>



Cigar String

Op	BAM	Description
M	0	alignment match (can be a sequence match or mismatch)
I	1	insertion to the reference
D	2	deletion from the reference
N	3	skipped region from the reference
S	4	soft clipping (clipped sequences present in SEQ)
H	5	hard clipping (clipped sequences NOT present in SEQ)
P	6	padding (silent deletion from padded reference)
=	7	sequence match
X	8	sequence mismatch



1. How many reads are in the tophat output file?

```
grep -v "^@SQ" accepted_hits.sam | wc
```

2. How many reads map to chromosome 4?

```
awk '$3=="SL2.40ch04" {print $0}' accepted_hits.sam | wc
```

3. How many reads map to each chromosome?

```
cut -f3 accepted_hits.sam | sort | uniq -c
```





- zero-based coordinates
- <http://genome.ucsc.edu/FAQ/FAQformat#format1>

chr7	127471196	127472363	Pos1	0	+	127471196	127472363	255,0,0
chr7	127472363	127473530	Pos2	0	+	127472363	127473530	255,0,0
chr7	127473530	127474697	Pos3	0	+	127473530	127474697	255,0,0
chr7	127474697	127475864	Pos4	0	+	127474697	127475864	255,0,0
chr7	127475864	127477031	Neg1	0	-	127475864	127477031	0,0,255
chr7	127477031	127478198	Neg2	0	-	127477031	127478198	0,0,255
chr7	127478198	127479365	Neg3	0	-	127478198	127479365	0,0,255
chr7	127479365	127480532	Pos5	0	+	127479365	127480532	255,0,0
chr7	127480532	127481699	Neg4	0	-	127480532	127481699	0,0,255



Popular file format for storing annotation information

- Column 1: "seqid"
- Column 2: "source" ex: software used
- Column 3: "type"
- Columns 4 & 5: "start" and "end"
- Column 6: "score" ex: e-values
- Column 7: "strand"
- Column 8: "phase" deals with where feature begins in ref to the reading frame
- Column 9: "attributes"

1-based coordinates

```
##gff-version 3
ctg123 . exon 1300 1500 . + . ID=exon00001
ctg123 . exon 1050 1500 . + . ID=exon00002
ctg123 . exon 3000 3902 . + . ID=exon00003
ctg123 . exon 5000 5500 . + . ID=exon00004
ctg123 . exon 7000 9000 . + . ID=exon00005
```



● Samtools tview

A terminal window titled "Terminal — samtools — 80x24" displaying the output of the samtools tview command. The output shows a genomic alignment track. The top line is the reference sequence: CAAGTCTCTTATGAATTAACCCAGTCAGACAAAAATAAGAAAAA...AATTTTAAAAATGAACAGAGCTTCAAGAAGTA. Below this, there are several lines of alignment data, including a line with a green 'R' and a line with a green 'A'. The alignment data is presented in a tabular format with columns for position, quality, and sequence. The terminal window has a black background and white text.



How good is the assembly?

- ❖ Check for contaminants by using BLAST to search appropriate databases (SeqClean)
- ❖ How many contigs are there vs how many genes expected - total sequence length?
- ❖ Length of contigs - compare to known long transcripts?
- ❖ How many reads map (reference-guided)?





Illumina

- ❖ Differential gene and transcript expression analysis of RNA-seq experiments with TopHat and Cufflinks (Trapnell et al., 2012)

454

- ❖ The comparison of gene expression from multiple cDNA libraries. (Stekel and Falciani, 2000).



- ◆ Expression experimental design - replicates, treatments, tissue, etc

Statistical Design and Analysis of RNA Sequencing Data
Paul L. Auer **and** R. W. Doerge **1**

- ◆ map to genome or transcriptome, is a reference genome available?





3. Analysis

Expression

Differential gene and transcript expression analysis of RNA-seq experiments with TopHat and Cufflinks

- [Cole Trapnell](#), [Adam Roberts](#), [Loyal Goff](#), [Geo Pertea](#), [Daehwan Kim](#), [David R Kelley](#), [Harold Pimentel](#), [Steven L Salzberg](#), [John L Rinn](#) & [Lior Pachter](#)

Bowtie
Extremely fast, general purpose short read aligner

TopHat
Aligns RNA-Seq reads to the genome using Bowtie
Discovers splice sites

Cufflinks package

Cufflinks
Assembles transcripts

Cuffcompare
Compares transcript assemblies to annotation

Cuffmerge
Merges two or more transcript assemblies

Cuffdiff
Finds differentially expressed genes and transcripts
Detects differential splicing and promoter use

CummeRbund
Plots abundance and differential expression results from Cuffdiff



RPKM

Reads Per Kilobase of exon model per Million mapped reads
(Mortazavi et al., 2008)

* Cufflinks uses the analogous FPKM

Computational methods for transcriptome annotation and quantification using RNA-seq . Garber et al., Nature Methods 2011





Exercise 2

Using the known tomato gene models for chromosome 4, detect differentially expressed gene in immature and breaker fruit.

STEP 1: Use cuffdiff for detection of differential expression.

```
cuffdiff -o cuffdiff_out -b ~/Desktop/ch4_demo_dataset/bwt2_index/SL2.40ch04.fa -u  
annotation/ITAG2.3_gene_models_ch4.gtf breaker/SRR404334/SRR404334_tophat_out/  
accepted_hits.bam,breaker/SRR404336/SRR404336_tophat_out/accepted_hits.bam  
immature_fruit/SRR404331/SRR404331_tophat_out/accepted_hits.bam,immature_fruit/  
SRR404333/SRR404333_tophat_out/accepted_hits.bam
```

~12 minutes





Expression

Cufflinks Output:

```
-rw-r--r-- 1 bioinfo bioinfo 12 Apr 16 11:17 cds.count_tracking
-rw-r--r-- 1 bioinfo bioinfo 115 Apr 16 11:17 cds.diff
-rw-r--r-- 1 bioinfo bioinfo 124 Apr 16 11:17 cds_exp.diff
-rw-r--r-- 1 bioinfo bioinfo 91 Apr 16 11:17 cds.fpkm_tracking
-rw-r--r-- 1 bioinfo bioinfo 115 Apr 16 11:17 cds.read_group_tracking
-rw-r--r-- 1 bioinfo bioinfo 343K Apr 16 11:17 gene_exp.diff
-rw-r--r-- 1 bioinfo bioinfo 183K Apr 16 11:17 genes.count_tracking
-rw-r--r-- 1 bioinfo bioinfo 333K Apr 16 11:17 genes.fpkm_tracking
-rw-r--r-- 1 bioinfo bioinfo 564K Apr 16 11:17 genes.read_group_tracking
-rw-r--r-- 1 bioinfo bioinfo 348K Apr 16 11:17 isoform_exp.diff
-rw-r--r-- 1 bioinfo bioinfo 189K Apr 16 11:17 isoforms.count_tracking
-rw-r--r-- 1 bioinfo bioinfo 346K Apr 16 11:17 isoforms.fpkm_tracking
-rw-r--r-- 1 bioinfo bioinfo 586K Apr 16 11:17 isoforms.read_group_tracking
-rw-r--r-- 1 bioinfo bioinfo 115 Apr 16 11:17 promoters.diff
-rw-r--r-- 1 bioinfo bioinfo 466 Apr 16 11:17 read_groups.info
-rw-r--r-- 1 bioinfo bioinfo 451 Apr 16 11:17 run.info
-rw-r--r-- 1 bioinfo bioinfo 115 Apr 16 11:17 splicing.diff
-rw-r--r-- 1 bioinfo bioinfo 124 Apr 16 11:17 tss_group_exp.diff
-rw-r--r-- 1 bioinfo bioinfo 12 Apr 16 11:17 tss_groups.count_tracking
-rw-r--r-- 1 bioinfo bioinfo 91 Apr 16 11:17 tss_groups.fpkm_tracking
-rw-r--r-- 1 bioinfo bioinfo 115 Apr 16 11:17 tss_groups.read_group_tracking
```

extract genes that have a significant value for differential expression

```
awk -F "\t" 'BEGIN {OFS = "\t"} $14 = "yes" {print $0}' gene_exp.diff >
significant_genes.txt
```




Expression

Cufflinks can also be used for:

- ◆ strand-specific RNA-seq
- ◆ novel transcript discovery in annotated genomes
- ◆ identification of novel splice variants
- ◆ detecting transcripts in genomes without annotation

For protocols see:

Differential gene and transcript expression analysis of RNA-seq experiments with TopHat and Cufflinks (Trapnell et al., 2012).