



Using R for GO Terms Analysis

sol genomics network

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by

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Using R for GO Terms Analysis:

1. What are GO terms ?
2. What is R ?
3. What is Bioconductor ?
4. Bioconductor modules for GO terms
 - 4.1. Basics: GO.db
 - 4.2. Gene Enrichment Analysis: TopGO
 - 4.3. GO profiles: GOProfiles
 - 4.4. Gene Similarities: GOSim
5. Example.
 - 5.1. Comparing two arabidopsis chromosomes





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I. What are GO Terms ?



Gene ontologies:

Structured controlled vocabularies (ontologies) that describe **gene products** in terms of their associated

biological processes,

cellular components and

molecular functions

in a species-independent manner

Ontology Statistics

As of ontology version 1.2279,
dated 23:09:2011 13:54

35017 terms, 100.0% defined

21432 biological_process

2897 cellular_component

9103 molecular_function

1585 obsolete terms (not
included in figures above)

<http://www.geneontology.org/GO.doc.shtml>





I. What are GO Terms ?



Biological processes,

Recognized series of events or molecular functions. A process is a collection of molecular events with a defined beginning and end.

Cellular components,

Describes locations, at the levels of subcellular structures and macromolecular complexes.

Molecular functions

Describes activities, such as catalytic or binding activities, that occur at the molecular level.





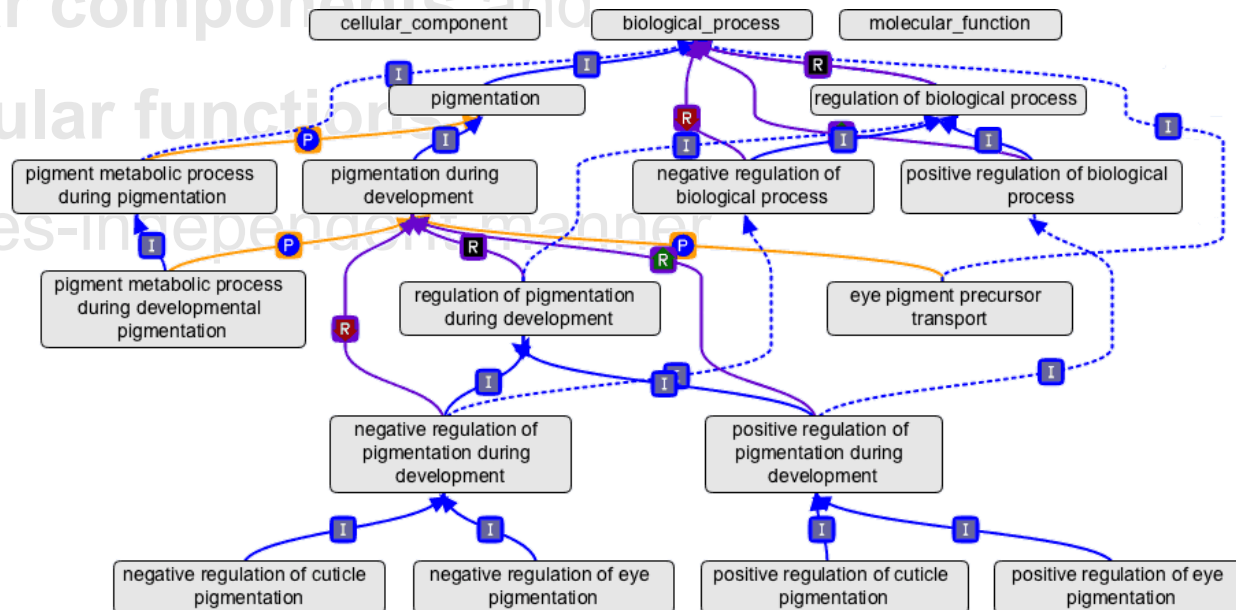
I. What are GO Terms ?



Gene ontologies:

Structured → **relationships between the terms**

GO can be described in terms of a **acyclic graph**, where:
each **GO term** is a **node**, and the
relationships between the terms are **arcs** between the nodes





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2. What is R ?

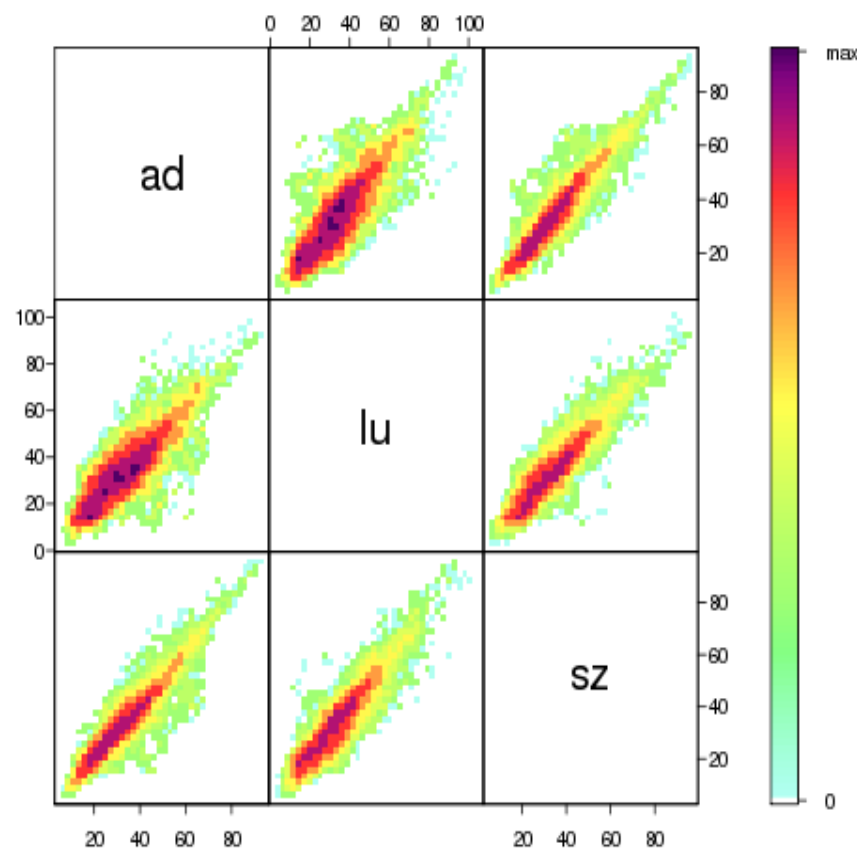


R is a **language** and **environment** for statistical computing and graphics.

```
## load required packages
library(IDPmisc)
library(SwissAir) # data for the example

## prepare the data
Ox <- AirQual[,c("ad.O3", "lu.O3", "sz.O3")] +
  AirQual[,c("ad.NOx", "lu.NOx", "sz.NOx")] -
  AirQual[,c("ad.NO", "lu.NO", "sz.NO")]
names(Ox) <- c("ad", "lu", "sz")

## draw graph
ipairs(Ox, ztransf = function(x){x[x<1] <- 1; log2(x)})
```





2. What is R ?



WEB:

OFICIAL WEB: <http://www.r-project.org/index.html>

QUICK-R: <http://www.statmethods.net/index.html>

BOOKS:

Introductory Statistics with R (Statistics and Computing), P. Dalgaard
[available as manual at R project web]

The R Book, MJ. Crawley

R itself:

`help()` and `example()`





2. What is R ?



INTEGRATED DEVELOPMENT ENVIRONMENT (IDE):

R-Studio: <http://rstudio.org/>

The screenshot shows the RStudio IDE interface. The top-left pane displays a data table with 11 rows and 4 columns (V1, V2, V3, V4). The top-right pane shows the Workspace with a 5x5 integer matrix 'mtx' and two integer vectors 'x' and 'y'. The bottom-left pane is the Console, showing the R version (2.12.1) and various help messages. The bottom-right pane is the Help pane, displaying the documentation for the 'na.fail' function, including its description, usage, and arguments.

	V1	V2	V3	V4
1	0	0	145	0
2	147	147	147	147
3	154	0	154	NA
4	0	0	221	NA
5	0	360	0	NA
6	445	445	445	NA
7	0	450	0	NA
8	0	458	0	NA
9	0	509	0	NA
10	0	510	0	NA
11	0	0	552	NA

```
R version 2.12.1 (2010-12-16)
Copyright (C) 2010 The R Foundation for Statistical Computing
ISBN 3-900051-07-0
Platform: x86_64-apple-darwin9.8.0/x86_64 (64-bit)

R is free software and comes with ABSOLUTELY NO WARRANTY.
You are welcome to redistribute it under certain conditions.
Type 'license()' or 'licence()' for distribution details.

Natural language support but running in an English locale

R is a collaborative project with many contributors.
Type 'contributors()' for more information and
'citation()' on how to cite R or R packages in publications.

Type 'demo()' for some demos, 'help()' for on-line help, or
'help.start()' for an HTML browser interface to help.
Type 'q()' to quit R.

[Workspace loaded from ~/.RData]
>
```

Handle Missing Values in Objects

Description

These generic functions are useful for dealing with `NA`s in e.g., data frames. `na.fail` returns the object if it does not contain any missing values, and signals an error otherwise. `na.omit` returns the object with incomplete cases removed. `na.pass` returns the object unchanged.

Usage

```
na.fail(object, ...)
na.omit(object, ...)
na.exclude(object, ...)
na.pass(object, ...)
```

Arguments



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3. What is Bioconductor ?



Bioconductor is an open source, open development software project to provide **tools** for the analysis and comprehension of **high-throughput genomic data**. It is based primarily on the **R programming language**.

Use Bioconductor for...



Microarrays

Import Affymetrix, Illumina, Nimblegen, Agilent, and other platforms. Perform quality assessment, normalization, differential expression, clustering, classification, gene set enrichment, genetical genomics and other workflows for expression, exon, copy number, SNP, methylation and other assays. Access GEO, ArrayExpress, Biomart, UCSC, and other community resources.



High Throughput Assays

Import, transform, edit, analyze and visualize flow cytometric, mass spec, HTqPCR, cell-based, and other assays.



Sequence Data

Import fasta, fastq, ELAND, MAQ, BWA, Bowtie, BAM, gff, bed, wig, and other sequence formats. Trim, transform, align, and manipulate sequences. Perform quality assessment, ChIP-seq, differential expression, RNA-seq, and other workflows. Access the Sequence Read Archive.



Annotation

Use microarray probe, gene, pathway, gene ontology, homology and other annotations. Access GO, KEGG, NCBI, Biomart, UCSC, vendor, and other sources.





3. What is Bioconductor ?

WEB:

OFFICIAL WEB: <http://www.bioconductor.org/>

BOOKS:

Bioinformatics and Computational Biology Solutions Using R and Bioconductor, 2005, Gentleman R. et al. (Springer)

R Programming for Bioinformatics, 2008, Gentleman R. (CRC Press)

MAILING LIST:

<http://www.bioconductor.org/help/mailing-list/>





3. What is Bioconductor ?



Installation of Bioconductor:

Use the `biocLite.R` script to install Bioconductor packages. To install a particular package, e.g., `limma`, type the following in an R command window:

```
source("http://bioconductor.org/biocLite.R")
biocLite("limma")
```

After downloading and installing this package, the script prints "Installation complete" and "TRUE". Install several packages, e.g., "GenomicFeatures" and "AnnotationDbi", with

```
biocLite(c("GenomicFeatures", "AnnotationDbi"))
```

To install a selection of core Bioconductor packages, use

```
biocLite()
```





3. What is Bioconductor ?



Use of Bioconductor:

Follow [installation instructions](#) to start using these packages. To install the annotations associated with the Affymetrix Human Genome U95 V 2.0, and with Gene Ontology, use

```
> source("http://bioconductor.org/biocLite.R")  
> biocLite(c("hgu95av2.db", "GO.db"))
```

Package installation is required only once per R installation. View a full list of available [software](#) and [annotation](#) packages.

To use the `AnnotationDbi` and `GO.db` package, evaluate the commands

```
> library("AnnotationDbi")  
> library("GO.db")
```

These commands are required once in each R session.





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4. Bioconductor modules for GO terms



Bioconductor Packages for GO Terms:

Bioconductor version 2.9 (Development)

- ▶ AnnotationData (604)
- ▶ ExperimentData (85)
- ▼ Software (499)
 - ▼ Annotation (66)
 - GO (15)
 - Pathways (23)
 - ProprietaryPlatforms (1)
 - ReportWriting (14)
 - ▶ AssayDomains (190)
 - ▶ AssayTechnologies (310)
 - ▶ Bioinformatics (290)
 - ▶ BiologicalDomains (50)
 - ▶ Infrastructure (201)

Packages

Software > Annotation > GO

- [annaffy](#) ▪ [annotate](#) ▪ [Category](#) ▪ [clusterProfiler](#) ▪ [gage](#) ▪ [globaltest](#) ▪ [GOFunction](#) ▪ [goProfiles](#)
- [GOSemSim](#) ▪ [goseq](#) ▪ [GOstats](#) ▪ [goTools](#) ▪ [mqsa](#) ▪ [RamiGO](#) ▪ [RNAither](#)





Bioconductor Packages for GO Terms:

[GO.db](#)

A set of annotation maps describing the entire Gene Ontology

[Gostats](#)

Tools for manipulating GO and microarrays

[GOSim](#)

functional similarities between GO terms and gene products

[GOProfiles](#)

Statistical analysis of functional profiles

[TopGO](#)

Enrichment analysis for Gene Ontology





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GO.db <http://www.bioconductor.org/packages/2.9/data/annotation/html/GO.db.html>

A set of annotation maps describing the entire Gene Ontology

1) GO terms stored in **objects**

GOTERM,

GOBPPARENTS, GOCCPARENTS, GOMFPARENTS

GOBPANCESTOR, GOCCANCESTOR, GOMFANCESTOR

GOBPCHILDREN, GOCCCHILDREN, GOMFCHILDREN

GOBPOFFSPRING, GOCCOFFSPRING, GOMFOFFSPRING

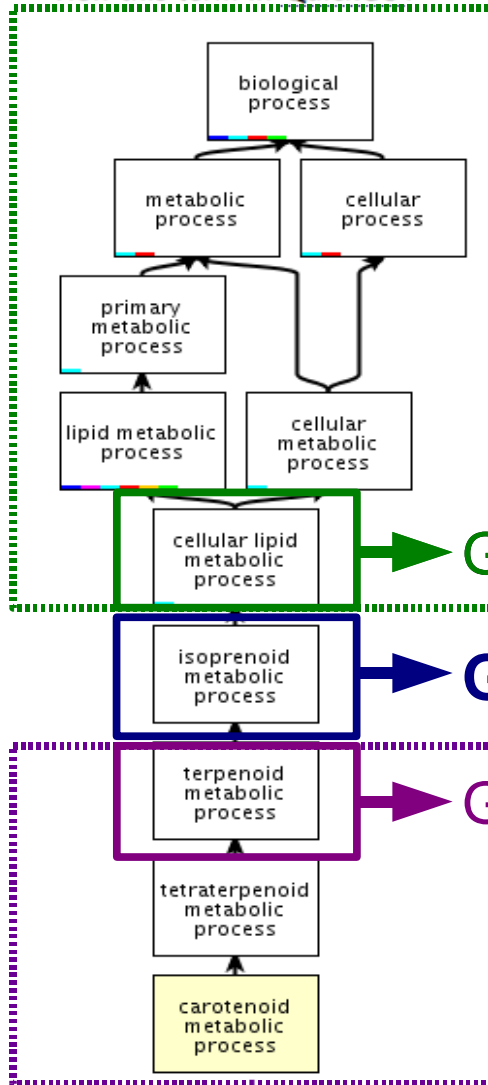
More Information:

<http://www.bioconductor.org/packages/2.6/bioc/vignettes/annotate/inst/doc/GOusage.pdf>





View this term in QuickGO.



GOBPANCESTOR → GOBPANCESTOR\$"GO:0006720"

GOBPPARENTS → GOBPPARENTS\$"GO:0006720"

GOTERM → GOTERM\$"GO:0006720"

GOBPCHILDREN → GOBPCHILDREN\$"GO:0006720"

GOBPOFFSPRING → GOBPOFFSPRING\$"GO:0006720"

QuickGO - <http://www.ebi.ac.uk/QuickGO>



Exercise 1:

- 1.1 Which term correspond top the GOID: GO:0006720 ?
- 1.2 How many synonyms has this term ?
- 1.3 How many children ?
- 1.4 ... and parents ?
- 1.5 ... and offsprings ?
- 1.6 ... and ancestors ?



2) Mapping between gene and GO terms stored in **objects** or **dataframes**.

Bioconductor version 2.8 (Release)

- ▼ AnnotationData (593)
 - ▶ ChipManufacturer (356)
 - ▶ ChipName (190)
 - CustomArray (2)
 - ▶ CustomCDF (16)
 - ▶ CustomDBSchema (10)
 - FunctionalAnnotation (10)
 - ▼ Organism (424)
 - Anopheles_gambiae (4)
 - Apis_mellifera (2)
 - Arabidopsis_thaliana (11)

Packages

AnnotationData > Organism > Arabidopsis_thaliana

- [aq.db](#) ▪ [aqcdf](#) ▪ [aqprobe](#) ▪ [arabidopsis.db0](#) ▪ [ath1121501.db](#) ▪ [ath1121501cdf](#) ▪ [ath1121501probe](#)
- [BSgenome.Athaliana.TAIR.04232008](#) ▪ [BSgenome.Athaliana.TAIR.TAIR9](#) ▪ [hom.At.inp.db](#) ▪ [org.At.tair.db](#)

org.At.tair.db

Genome wide annotation for Arabidopsis, primarily based on mapping using TAIR identifiers.



2) Mapping between gene and GO terms stored in **objects** (**annotate package**) or **dataframes**.

```
> library("org.At.tair.db")  
> org.At.tairGO[["AT5G58560"]]
```


Use a list:

```
> org.At.tairGO[["AT5G58560"]][[1]]$Ontology
```

Functions (“annotate” package):

- + getOntology(inlist, gocategorylist)
- + getEvidence(inlist)

```
> getOntology(org.At.tairGO[["AT5G58560"]])  
> getEvidence(org.At.tairGO[["AT5G58560"]])
```



Exercise 2:

- 2.1 How many terms map with the gene: AT5G58560 ?
- 2.2 What biological process terms map with this gene?
- 2.3 What evidences are associated with these terms ?





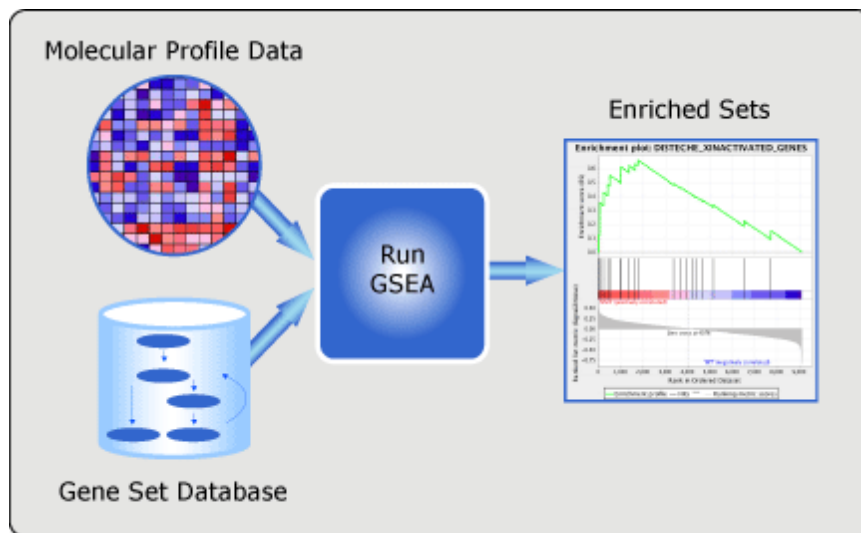
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Gene Set Enrichment Analysis (GSEA)

Computational method that determines whether an a priori defined set of genes shows statistically significant, concordant differences between two biological states (e.g. phenotypes).



c1 **positional gene sets** for each human chromosome and each cytogenetic band.

c2 **curated gene sets** from online pathway databases, publications in PubMed, and knowledge of domain experts.

c3 **motif gene sets** based on conserved *cis*-regulatory motifs from a comparative analysis of the human, mouse, rat and dog genomes.

c4 **computational gene sets** defined by expression neighborhoods centered on 380 cancer-associated genes.

c5 **GO gene sets** consist of genes annotated by the same GO terms.

topGO <http://www.bioconductor.org/packages/release/bioc/html/topGO.html>

topGO: Enrichment analysis for Gene Ontology

	fisher	ks	t	globaltest	sum
classic	✓	✓	✓	✓	✓
elim	✓	✓	✓	✓	✓
weight	✓	—	—	—	—
weight01	✓	✓	✓	✓	✓
lea	✓	✓	✓	✓	✓
parentchild	✓	—	—	—	—

Table 1: *Algorithms currently supported by topGO.*



Methodology:

1) Data preparation:

- a) Gene Universe.
- b) GO Annotation.
- c) Criteria to select interesting genes.

OBJECT:
topGOdata

```
> sampleGOdata <- new("topGOdata",  
  description = "Simple session",  
  ontology = "BP",  
  allGenes = geneList,  
  geneSel = topDiffGenes,  
  nodeSize = 10,  
  annot = annFUN.db,  
  affyLib = "hgu95av2.db")
```

Gene Universe

Selected Genes

Function to map data provided
in the annotation data packages

Data package



```
> sampleGOdata
```

```
----- topGOdata object -----
```

Description:

- Simple session

Ontology:

- BP

323 available genes (all genes from the array):

- symbol: 1095_s_at 1130_at 1196_at 1329_s_at 1340_s_at ...
- score : 1 1 0.62238 0.541224 1 ...
- 50 significant genes.

316 feasible genes (genes that can be used in the analysis):

- symbol: 1095_s_at 1130_at 1196_at 1329_s_at 1340_s_at ...
- score : 1 1 0.62238 0.541224 1 ...
- 50 significant genes.

GO graph (nodes with at least 10 genes):

- a graph with directed edges
- number of nodes = 672
- number of edges = 1358

```
----- topGOdata object -----
```

OBJECT:
topGOdata



Methodology:

2) Running the enrichment test: (**runTest** function)

```
> resultFisher <- runTest(sampleGOdata,  
  algorithm = "classic",  
  statistic = "fisher")
```

```
> resultFisher
```

Description: Simple session

Ontology: BP

'classic' algorithm with the 'fisher' test

672 GO terms scored: 66 terms with $p < 0.01$

Annotation data:

Annotated genes: 316

Significant genes: 50

Min. no. of genes annotated to a GO: 10

Nontrivial nodes: 607

whichAlgorithms()

whichTest()

"fisher"

"ks"

"t"

"globaltest"

"sum"

"ks.ties"



Methodology:

2) Running the enrichment test:

```
> resultKS <- runTest(sampleGOdata,  
  algorithm = "classic",  
  statistic = "ks")
```

```
> resultKS
```

Description: Simple session

Ontology: BP

'classic' algorithm with the 'ks' test

672 GO terms scored: 57 terms with $p < 0.01$

Annotation data:

Annotated genes: 316

Significant genes: 50

Min. no. of genes annotated to a GO: 10

Nontrivial nodes: 672

whichAlgorithms()

whichTest()

"fisher"

"ks"

"t"

"globaltest"

"sum"

"ks.ties"



Methodology:

3) Analysis of the results: (**GenTable** function)

```
> allRes <- GenTable(sampleGOdata,  
  classicFisher = resultFisher,  
  classicKS = resultKS,  
  orderBy = "classicKS",  
  ranksOf = "classicFisher",  
  topNodes = 10)
```

```
> allRes
```

	GO.ID	Term	Annotated	Significant	Expected	Rank in classicFisher	classicFisher	classicKS
1	GO:0007049	cell cycle	204	33	32.28	324	0.48	2.8e-11
2	GO:0022403	cell cycle phase	189	28	29.91	435	0.78	8.2e-11
3	GO:0022402	cell cycle process	192	29	30.38	423	0.73	1.5e-10
4	GO:0000278	mitotic cell cycle	191	28	30.22	442	0.81	2.0e-10
5	GO:0000087	M phase of mitotic cell cycle	185	24	29.27	566	0.96	2.9e-10
6	GO:0000279	M phase	185	24	29.27	567	0.96	2.9e-10
7	GO:0000280	nuclear division	176	18	27.85	601	1.00	3.7e-09
8	GO:0007067	mitosis	176	18	27.85	602	1.00	3.7e-09
9	GO:0048285	organelle fission	176	18	27.85	603	1.00	3.7e-09
10	GO:0006996	organelle organization	190	21	30.06	600	1.00	2.1e-08





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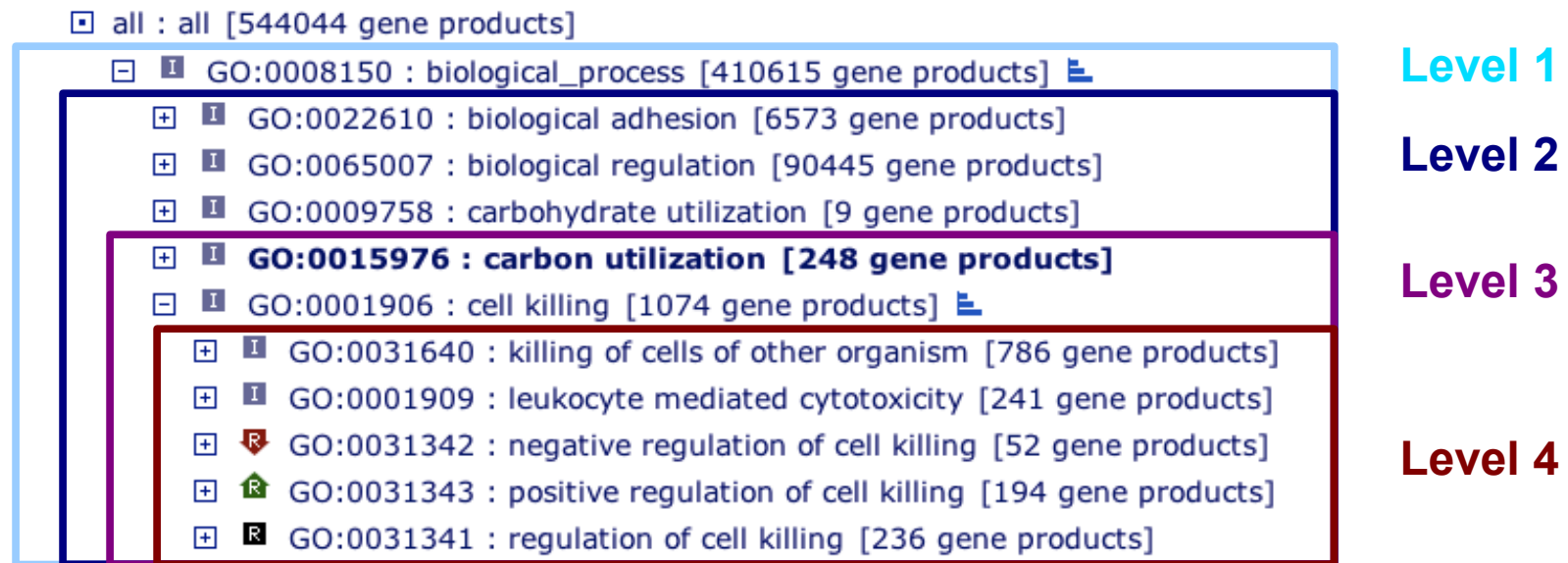
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goProfiles <http://www.bioconductor.org/packages/2.8/bioc/html/goProfiles.html>

goProfiles: an R package for the statistical analysis of functional profiles

1) Profiles are built by slicing the GO graph at a given level





4.3 goProfiles



2) Functional profile at a given GO level is obtained by counting the number of identifiers having a hit in each category of this level

<i>GO Term</i>	<i>gene1</i>	<i>gene2</i>	<i>gene3</i>	<i>gene4</i>
GO:0005488	1	1	0	0
GO:0030234	0	1	1	1
GO:0045182	0	0	0	1

Table 2.1: A simple list of 4 identifiers considered at level 2 of the MF ontology illustrates how some genes may have hits in several categories simultaneously.

3) Profiles comparissons:

1. How different or representative of a given gene universe is a given set of genes?
 - Universe: All genes analyzed,
Gene Set: Differentially expressed genes in a microarray experiment
 - Universe: All genes in a database,
Gene Set: Arbitrarily selected set of genes
2. How biologically different are two given sets of genes?
 - Differentially expressed genes in two experiments
 - Arbitrarily chosen lists of genes





4.3 goProfiles



Methodology:

1) Data preparation:

a) Gene GO term map (**object** or **dataframe**).

Dataframe with 4 columns:

- + GeneID
- + Ontology
- + Evidence
- + GOID

2) Profile creation:

a) **basicProfile** function

```
BasicProfile( genelist,  
              idType = "Entrez",  
              onto = "ANY",  
              Level = 2,  
              orgPackage=NULL,  
              anotPackage=NULL,  
              ...)
```

“Entrez” (default),
“BiocProbes”,
“GoTermsFrame”

Requested for “Entrez”

Requested for
“BiocProbes”





4.3 goProfiles



Methodology:

2) Profile creation:

a) **basicProfile** function

```
> printProfiles(bpprofile)
```

```
Functional Profile
```

```
=====
```

```
[1] "BP ontology"
```

	Description	GOID	Frequency
25	cellular component biogen...	GO:0044085	5
14	cellular component organi...	GO:0016043	6
12	cellular process	GO:0009987	122
32	establishment of localiza...	GO:0051234	4
4	immune system process...	GO:0002376	1
31	localization	GO:0051179	4
2	metabolic process	GO:0008152	3
33	multi-organism process...	GO:0051704	2
30	response to stimulus	GO:0050896	4





4.3 goProfiles



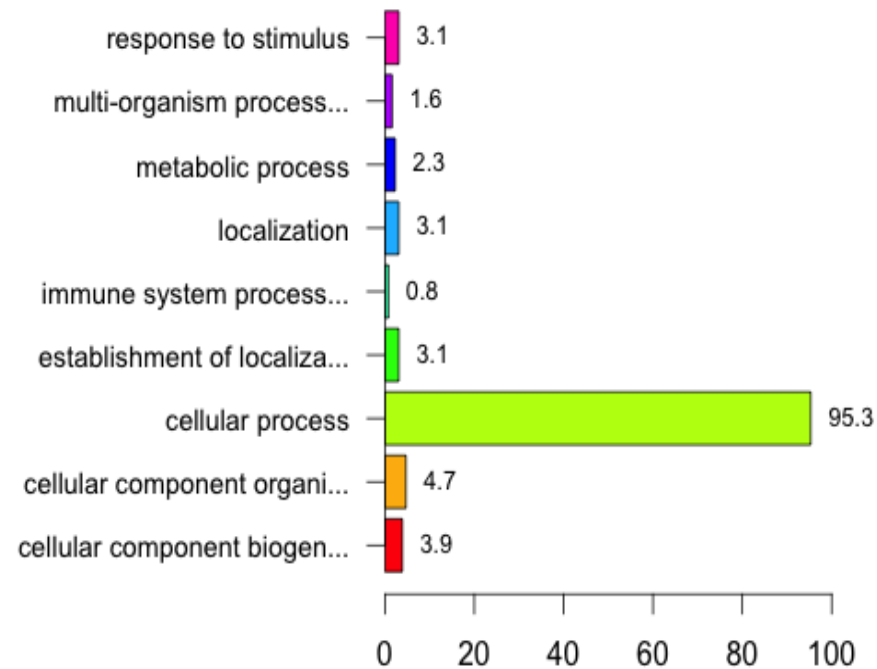
Methodology:

2) Profile creation:

a) **basicProfile** function

```
> plotProfiles(bpprofile)
```

Functional Profile. BP ontology





4.3 goProfiles



Methodology:

2) Profile creation:

b) **expandedProfile** function

Used mainly for comparisons of profiles.

```
ExpandedProfile( genelist,  
                 idType = "Entrez",  
                 onto = "ANY",  
                 Level = 2,  
                 orgPackage=NULL,  
                 anotPackage=NULL  
                 ...)
```

3) Profile comparisons:

- Case I (INCLUSION): One list included in the other.
- Case II (DISJOINT): Non overlapping gene sets
- Case III (INTERSECTION): Overlapping genes





4.3 goProfiles



Methodology:

3) Profile comparisons:

- Case I (INCLUSION): `compareProfilesLists()`
- Case II (DISJOINT): `compareGeneLists()`
- Case III (INTERSECTION): `compareGeneLists()`

```
> comp_ath_genes <- compareGeneLists(  
  genelist1=ath_chl_list,  
  genelist2=ath_mit_list,  
  idType="Entrez", orgPackage="org.At.tair.db",  
  onto="BP", level=2)
```

```
> print(compSummary(comp_ath_genes))
```

Sqr.Euc.Dist	StdErr	pValue	0.95Cl.low	0.95Cl.up
0.031401	0.024101	0.005000	-0.015837	0.078639





4.3 goProfiles



Methodology:

3) Profile comparisons:

```
> basic_mitprof <- basicProfile(ath_mit_list, idType="Entrez",  
  onto="BP", level=2, orgPackage="org.At.tair.db",  
  empty.cats=TRUE)  
  
> basic_chloprof <- basicProfile(ath_chl_list, idType="Entrez",  
  onto="BP", level=2, orgPackage="org.At.tair.db",  
  empty.cats=TRUE)  
  
> merged_prof <- mergeProfilesLists(basic_chloprof, basic_mitprof,  
  profNames=c("Chloroplast", "Mitochondria"))  
  
> plotProfiles(merged_prof, percentage=TRUE, legend=TRUE)
```





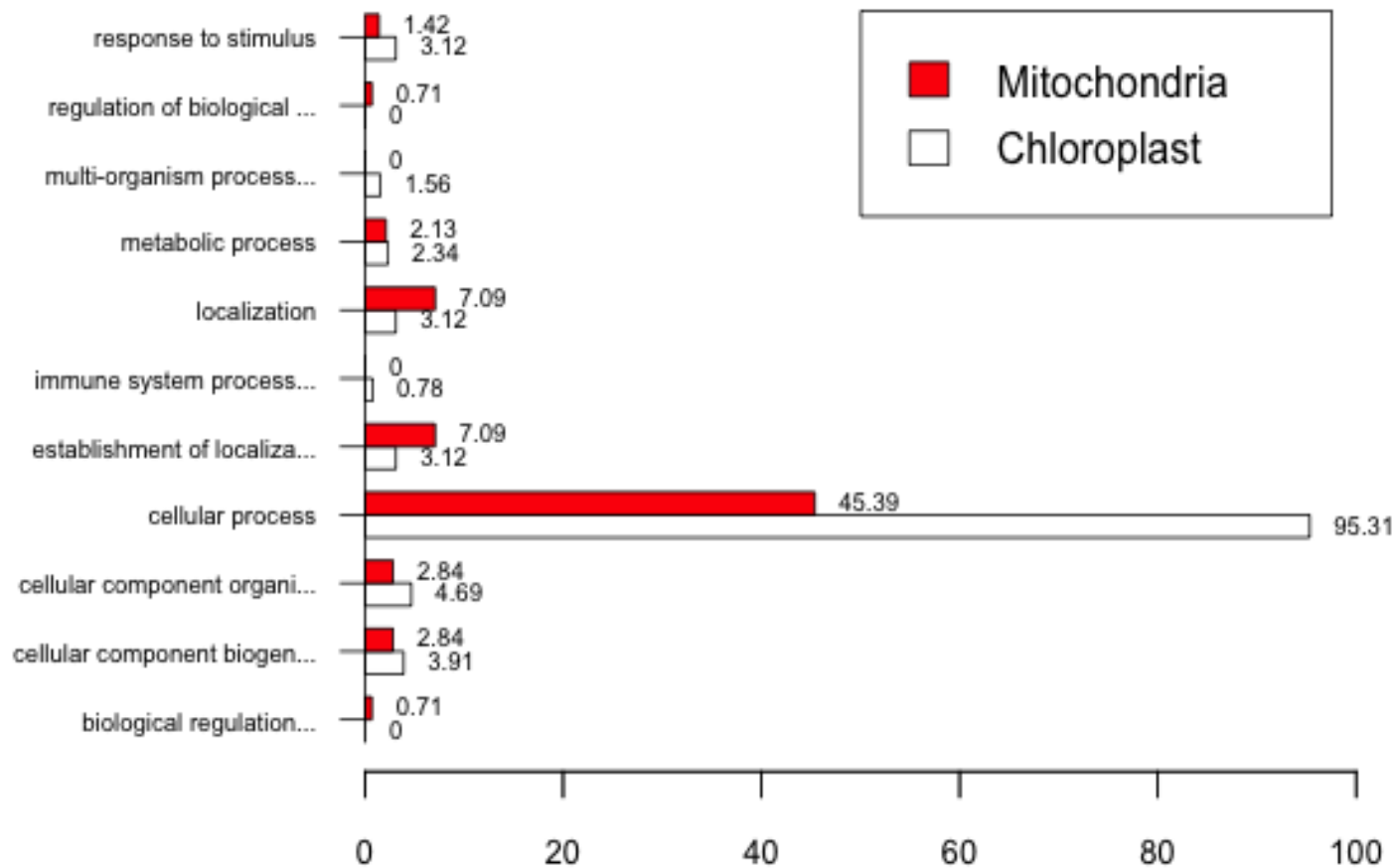
4.3 goProfiles



Methodology:

3) Profile comparisons:

Functional Profile. BP ontology





Using R for GO Terms Analysis:

1. What are GO terms ?
2. What is R ?
3. What is Bioconductor ?
4. Bioconductor modules for GO terms
 - 4.1. Basics: GO.db
 - 4.2. Gene Enrichment Analysis: TopGO
 - 4.3. GO profiles: GOProfiles
 - 4.4. Gene Similarities: GOSim**
5. Example.
 - 5.1. Comparing two arabidopsis chromosomes.





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Exercise 3:

Compare the goProfiles for *A. thaliana* chromosomes 1 and 5.

Are they significantly different ?

Source:

<ftp://ftp.arabidopsis.org/home/tair/Genes/>

[TAIR10_genome_release/TAIR10_gene_lists/TAIR10_all_gene_models](ftp://ftp.arabidopsis.org/home/tair/Genes/TAIR10_genome_release/TAIR10_gene_lists/TAIR10_all_gene_models)

R Package:

GoProfiles

